



SRF-RE Luciferase Reporter-HEK293 Cell Line

Catalog number: RC1033

This package insert must be read in its entirety before using this product.

For research use only. Not for use in diagnostic procedures.

SRF-RE Luciferase Reporter-HEK293 Cell Line



Contents: Each vial contains 2 ~ 3 x 10⁶ cells in 1 ml of 90% FBS + 10% DMSO.

Description: The SRF-RE Luciferase Reporter cell line is a stably transfected HEK 293 cell line which expresses Renilla luciferase reporter gene under the transcriptional control of the serum response factor-response element (SRF-RE). The SRF-RE reporter cell line can be used for studying GPCR-linked RhoA signaling pathways as well as screening of agonists, antagonists or signaling inhibitors related with the RhoA signaling pathways. Functional activity of the cell line has been validated by serum stimulation assay (Figure 1).

Applications: Functional Assay

Application Notes: Functional Assay, detecting the transcriptional activity of SRF-RE

Application Details: Dilute the sample so that the expected range of concentrations fall within the detection range of this kit.

If the expected range of concentration is unknown, a pilot test should be conducted to decide the optimal dilution ratio for your samples.

Some PubMed article(s) citing the expression level of this target are as follows:

Boster Bio's internal QC testing used:

Application:

Monitor the GPCR-linked RhoA signaling pathway. Screen for activators or inhibitors of the GPCR-linked RhoA signaling pathway.

Culture conditions:

Cells should be grown at 37°C with 5% CO2 using DMEM medium supplemented with 10% FBS and 1% Pen/Strep, plus 3 µg/ml of Puromycin. It is recommended to quickly thaw the frozen cells upon receipt or from liquid nitrogen in a 37°C water-bath, transfer to a tube containing 10 ml of growth medium without Puromycin, spin down cells, resuspend cells in pre-warmed growth medium without Puromycin, transfer resuspended cells to T25 flask and culture in 37°C-CO2 incubator. Leave the T25 flask in the incubator for 2~4 days without disturbing or changing the medium until cells completely recover viability and become adherent. Once cells are over 90% adherent, remove growth medium and passage the cells through trypsinization and centrifugation. At first passage, switch to growth medium containing Puromycin. Cells should be split before they reach complete confluence. To passage the cells, detach cells from culture vessel with Trypsin/EDTA, add complete growth medium and transfer to a tube, spin down cells, resuspend cells and seed appropriate aliquots of cells suspension into new culture vessels. Subcultivation ration = 1:10 to 1:20 weekly.

Functional validation:

A. Response of SRF-RE HEK293 cells to fetal bovine serum (FBS). 1. Harvest SRF-RE HEK293 cells and seed cells into a white solid-bottom 96-well microplate at 5×10^4 cells/well in $100 \, \mu$ l of growth medium containing 0.5% FBS to start serum starvation. 2. Incubate cells at

SRF-RE Luciferase Reporter-HEK293 Cell Line (RC1033) Images

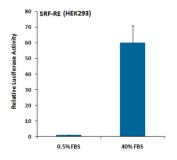


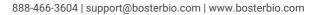
Fig-1: Induction of SRF-RE activity by FBS in SRF-RE HEK293 cells.

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